

Maternal Adaptation to High-altitude Pregnancy: An Experiment of Nature—A Review

L. G. Moore^{a,b,*}, M. Shriver^c, L. Bemis^b, B. Hickler^a, M. Wilson^a, T. Brutsaert^d, E. Parra^{c,e} and E. Vargas^f

^a Department of Anthropology, University of Colorado at Denver, Denver, CO, USA; ^b Colorado Center for Altitude Medicine and Physiology, University of Colorado Health Sciences Center, 4200 East Ninth Avenue, Denver, CO 80262, USA;

^c Department of Anthropology, Pennsylvania State University, State College, PA, USA; ^d Department of Anthropology, State University of New York (SUNY) at Albany, NY, USA; ^e Department of Anthropology, University of Toronto, Mississauga, Ontario, Canada; ^f Instituto Boliviano de Biología de Altura (Bolivian High-Altitude Biology Institute), La Paz, Bolivia

Paper accepted 2 January 2004

A long and productive history of studies at high altitude has demonstrated that chronic hypoxia plays a key role in the aetiology of intrauterine growth restriction (IUGR) and pre-eclampsia. Susceptibility to altitude-associated IUGR varies among high-altitude populations in relation to their duration of altitude exposure, with multigenerational residents demonstrating one-third the birth weight fall present in shorter-resident groups. Higher uteroplacental blood flow during pregnancy in multigenerational high-altitude residents suggests that such population differences are due, at least in part, to differences in maternal vascular responses to pregnancy. We hypothesize that natural selection acting on hypoxia-inducible factor (HIF)-targeted or -regulatory genes has enabled maternal vascular adaptation to pregnancy in long-resident high-altitude groups. Preliminary evidence in support of this hypothesis demonstrates that the potent HIF-targeted vasoconstrictor, endothelin-1 (ET-1), is differentially regulated by pregnancy and chronic hypoxia in Andean vs European residents of high altitude. Andeans show the normal, pregnancy-associated fall in ET-1 levels previously reported at low altitude, whereas Europeans have higher ET-1 levels and little pregnancy-associated change, like pre-eclamptic women. Single nucleotide polymorphisms (SNPs) in the ET-1 gene also differ in Andeans compared with low-altitude populations. We conclude that high altitude serves as an experiment of nature for elucidating genetic factors underlying susceptibility to complications of pregnancy and fetal life. Such studies may be important for identifying persons at risk for these complications at any altitude. © 2004 IFPA and Elsevier Ltd. All rights reserved.

Placenta (2004), 25, Supplement A, Trophoblast Research, Vol. 18, S60–S71

INTRODUCTION

Hypoxia is a frequent complication of prenatal life. When prolonged, it is associated with intrauterine growth restriction (IUGR) (Table 1) and increased perinatal mortality and morbidity. Whereas persons are subject to chronic hypoxia in utero at all elevations, the largest single group of persons at risk is the 140 million worldwide residents of high altitude (>2500 m or 8000 ft) [1]. There has been a long and productive history of studies of pregnancy, fetal and neonatal life at high altitude. The initial observations on a population level that fetal growth restriction and preterm delivery were separable causes of low birth weight were made there nearly 50 years ago ([2] reviewed in [3]). Not only was this crucial in terms of our current understanding of the causes of low birth weight but was followed by continued investigations demonstrating the utility of charting newborn birth weight in relation to gestational age as a predictor of infant mortality [4]. This system continues to

be used worldwide; its introduction constitutes one of the truly great public health advances of our time. The first recognition that chronic hypoxia was part of the aetiology of pre-eclampsia was also made at high altitude [5]. Continuing investigations by several investigative groups [6–9] are rapidly expanding our knowledge of the cellular and molecular mechanisms by which hypoxia influences the maternal, placental and fetal responses required to successfully produce the next generation.

Here we begin with a brief review of the magnitude and cause of the altitude-associated increase in IUGR. Because studies have been conducted at a range of elevations on three continents (North America, South America, Asia), we ask whether the decline in infant birth weight varies among populations and if so, what mechanisms are likely involved. We then move to a consideration of the physiological factors controlling uteroplacental blood flow since altered uteroplacental blood flow is a core predictor of pregnancy abnormalities [10]. After reviewing recent research addressing the effects of chronic hypoxia on uteroplacental blood flow, we ask whether these effects vary among populations or species in relation to

* To whom correspondence should be addressed. Fax: +1-303-315-4871; E-mail: lorna.g.moore@uchsc.edu

Table 1. Key to abbreviations

ARNT	ADP-ribosylation factor domain protein 1	PHD	Prolyl hydroxylase domain
EI	External iliac	PIGF	Placenta like growth factor
ET-1	Endothelin 1	sFlt-1	Membrane bound VEGF receptor 1
HIF	Hypoxia-inducible factor	SNP	Single nucleotide polymorphism
HRE	Hypoxia responsive element	SVR	Systemic vascular resistance
IUGR	Intrauterine growth restriction	UA	Uterine artery
LSBL	Locus-specific branch length	VEGF	Vascular endothelial growth factor
NO	Nitric oxide	VHL	von Hippel Lindau
NOS	Nitric oxide synthase		

the altitude-associated increase in IUGR. Finally, we consider the role that genetic factors may play in the population and species differences observed in altitude-associated IUGR, hypothesizing that variation in hypoxia-sensitive genes might be involved. We review the role played by the hypoxia-inducible factor (HIF) pathway since it is responsible for regulating most of the oxygen-sensitive genes. After surveying the HIF-regulated and regulatory genes differentially affected by pregnancy and chronic hypoxia, we consider the possibility that such genes may have been acted upon by natural selection in populations long resident at high altitude. We conclude with some future directions for research aimed at advancing our understanding of the genetic mechanisms regulating maternal physiological responses to human pregnancy.

ALTITUDE-ASSOCIATED IUGR

Magnitude and cause of the birth weight decline

In Colorado and elsewhere, infant birth weight declines with increasing altitude, averaging a 100 g fall per 1000 m altitude gain (nearly a quarter of a pound per 3000 ft) [11,12]. While convenient to express in this fashion, the decline in birth weight is actually curvilinear with the ‘breakpoint’ occurring about 2000 m or 6600 ft [13], consistent with the shape of the haemoglobin-oxygen dissociation curve. The entire distribution of birth weights is shifted to lower values, rather than only a subset of lower birth weight babies being affected [2]. This leftward shift has the net effect of increasing the proportion of low birth weight infants (<2500 g) by more than 50 per cent [11]. In Colorado, the effect of high altitude on birth weight is as great or greater than that associated with low maternal weight gain, smoking, primiparity or pre-eclampsia [11].

The decline in birth weight is principally due to a slowing of fetal growth in the 3rd trimester. Gestational age is, on average, 0.5 week shorter at elevations over 2744 m (9000 ft) in Colorado, but this is not sufficient to explain the 240 g birth weight fall [11]. Comparisons of weights of babies born prematurely suggest that fetal growth begins to slow after 28–31 weeks’ gestation [14]. Such findings have been recently confirmed by Krampfl and co-workers who found a reduction in fetal biometry dimensions from 25–29 weeks’ onwards, with

abdominal circumference being more affected than head circumference [15].

The incidence of pre-eclampsia is increased at high altitude and this likely contributes to the altitude-associated birth weight decline [5,11,16,17]. In fact, the greater incidence of pre-eclampsia accounts for about half the birth weight decline in our recent studies in Bolivia [17]. Consistent with the concept that not just a subset of women but the population of pregnant women is affected, even normotensive women in Colorado fail to show the normal pregnancy-associated blood pressure fall suggesting a generalized disorder in maternal vascular adjustment to pregnancy [16].

The increased incidence of IUGR and pre-eclampsia at high altitude likely contributes to a rise in perinatal and infant mortality and morbidity. In a large chart-review of women receiving prenatal care at similar kinds of health care facilities at low (300 m) vs high (3600 m) altitude in Bolivia, the combination of high-altitude residence and pre-eclampsia raised the frequency of stillbirths 3-fold [17]. Not only were IUGR and pre-eclampsia more common but all the other pregnancy, fetal and newborn complications surveyed were more frequent at the high- than the low-altitude site [17,18]. Infant mortality increases proportionally to the rise in IUGR in nearly all studies [2,19–23]. In the one report in which such an increase did not occur [14], women from high altitudes used specialized medical services to a greater extent than did the low-altitude residents. Thus, the increased incidence of IUGR and pre-eclampsia at high altitude appear to raise infant mortality. Whether a different standard should be applied for determining IUGR at high vs low altitude, as has been called for recently [15], awaits determination of the birth weight-specific infant mortality (and morbidity) risks at high altitude vs low altitude, with adequate controls for gestational age and other key factors such as medical care. Such an assessment is especially important for the developing regions of South America where infant mortality rates are among the highest in the western hemisphere [24] in order for the standard to be able to identify as many high-risk infants as possible.

Interpopulational variation in the altitude-associated birth weight decline

Population variation in the magnitude of the birth weight reduction at high altitude was first noted in Andean vs

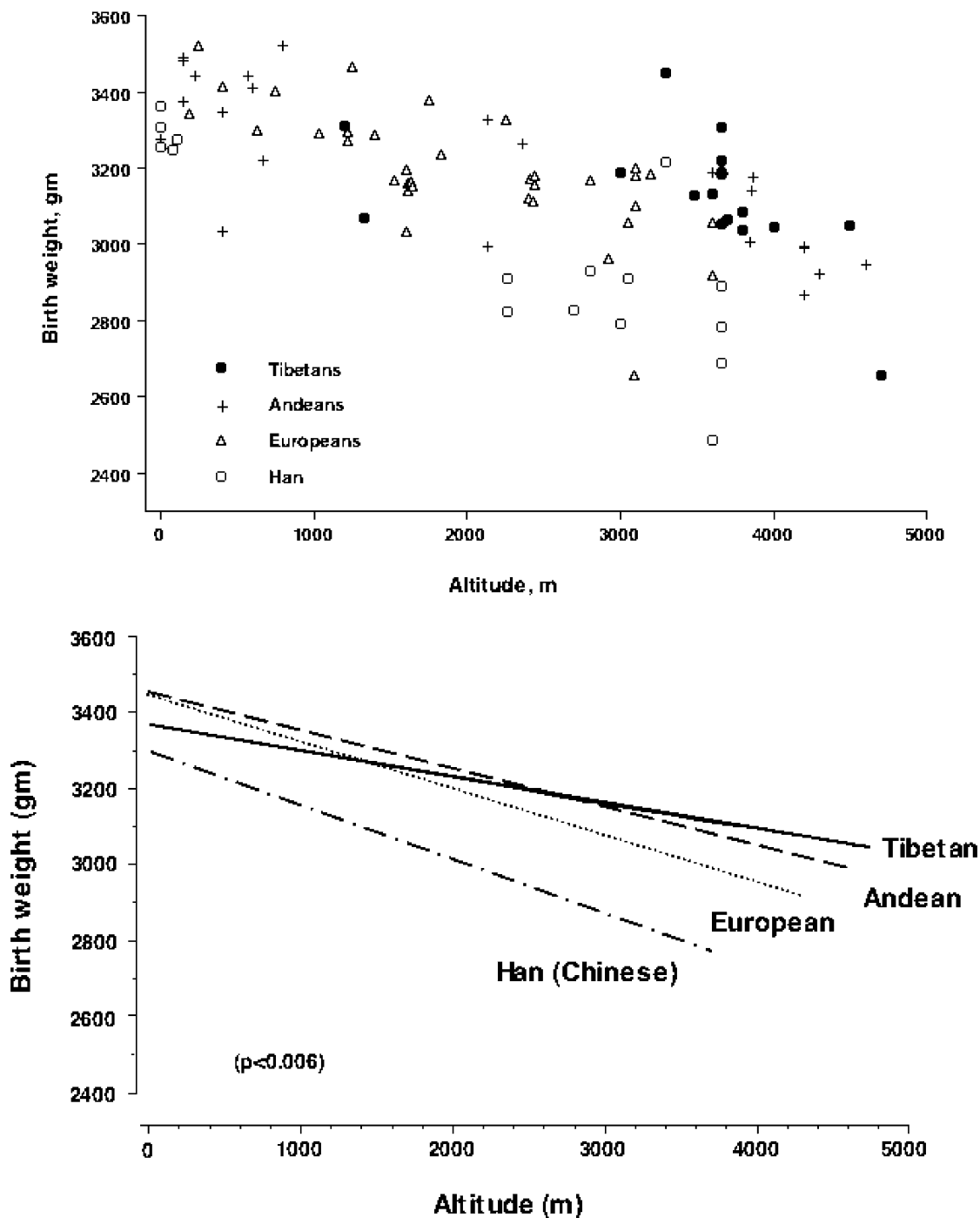


Figure 1. (a) Data points are average values for ~4 million births occurring in the populations and altitudes represented. The Tibetan population is likely to have lived the longest at high altitude, followed by Andeans, Europeans and then Han. For original data sources, see [26]. (b) Best fit regression lines for the data presented in 1a, weighted by sample size and variance, demonstrate that the magnitude of altitude-associated birth weight decline varies inversely with the duration (in generations) of altitude residence ($P < 0.0006$).

European women in La Paz, Bolivia (3600 m) [25]. In a large review of some 4 million births, we found that while the general pattern was for birth weight to decline, the birth weights observed at a given altitude were highly variable (Figure 1A) [26]. But when grouped by population ancestry (Figure 1B), a consistent pattern emerged.

Specifically, Tibetans and Andeans who have lived at high altitudes for 10 000 years (Andeans) to 20 000 years (Tibetans) show one-third the birth weight reduction present in European or Han ('Chinese') populations that have resided at high altitudes for <500 years (i.e. <400 years in South America, <150 years in North America, and ~ 50 years in western China

Table 2. Pre-natal and post-natal mortality (deaths per 1000 pregnancies or livebirths) in women of low vs high socioeconomic status living in La Paz, Bolivia (elevation 3600 m) evaluated at 2.67 gravidity and 2.20 parity

	Income group	Unadjusted means	Adjusted means	Mean difference	<i>F</i>	<i>P</i> value
Pre-natal mortality	High	120.0	137.0	-26.0	7.796	0.005
	Low	127.8	111.0			
<20 weeks	High	113.0	129.0	-34.3	14.8	0.000
	Low	109.7	94.3			
≥20 weeks	High	7.7	9.3	8.9	4.817	0.028
	Low	18.1	18.2			
Post-natal mortality	High	5.9	10.9	25.7	27.706	0.000
	Low	42.7	36.6			

[26]). Thus, for example, we found babies born to Tibetan mothers weighed 310 g more at 2700–3000 m (95 per cent CI=126, 494 g; $P<0.01$) and 530 g more at 3000–3800 m (210, 750 g; $P<0.01$) than babies born to Han mothers residing in the Tibet Autonomous Region of southwestern China. When viewed across their respective altitude ranges, Tibetans living at 2700–4800 m demonstrated a 15 g reduction in birthweight per 1000 m altitude gain whereas Han residing at 2700–3800 m had a 45 g/1000 m birthweight fall [27]. Such interpopulation variation does not appear attributable to differences in maternal body size, nutrition, or health care [25,26]. Since the Andean and Tibetan women are likely to have been born and raised at high altitude whereas the European and Han women may have moved there more recently, it is possible that some of these population differences are due to factors stemming from the woman's own altitude of birth and development. However, in our and others' Colorado studies, lifelong high-altitude residence does not appear to protect against altitude-associated IUGR [28,29]. It is likely that population differences in altitude-associated IUGR influence infant mortality. In the high-altitude regions of Tibet where Tibetan and Han populations live under conditions of similar and, in communist China, no-cost health care, not only were Tibetans protected from altitude-associated birthweight declines but they also had lower estimated pre- and post-natal mortality rates than the Han residing at the same altitudes [27]. Clearly, socioeconomic and health care characteristics are also key factors affecting infant mortality but these observations suggest that population-specific, genetic factors may also be involved.

In Bolivia we used surnames to compare the altitude-associated increase in IUGR among Andean, mestizo ('mixed'), and European surnamed segments of the population. Surnames as a means for assessing group-level population ancestry has been validated by comparison with genetic markers in the Bolivian population [30]. In approximately 1000 consecutive births to women receiving prenatal care and living at low (300 m, Santa Cruz), medium (2500 m, Cochabamba), or high (3600 m, La Paz and Oruro) altitude, we found that babies of Andean ancestry had only one-third as much altitude-associated increase in IUGR as babies of European ancestry [31]. Such findings help to explain why babies born to the higher-socioeconomic and more often European segment

of the population at high altitude weigh less than those of the Andean, often lower-socioeconomic sector [12].

To evaluate the effects of socioeconomic vs population-ancestry characteristics on birth weight and on pre- and post-natal survival in Bolivia, we compared 1602 deliveries to high-altitude (3600 m) residents from households with above average vs below average monthly incomes (>\$500 and <\$500/month, $n=817$ and 785 respectively). Similar to previous reports [12], the frequency of low birth weight (<2500 g) was greater in the high- than low-income households (10.3 vs 7.0 per cent, $P<0.02$). Using logistic regression to take into account the effects of maternal education (another socioeconomic indicator), high income increased the likelihood of having a low birth weight baby [OR=1.69 (1.08, 2.64) 95 per cent confidence intervals] whereas Andean ancestry reduced the risk [OR=0.35 (0.14, 0.86)]. We also estimated pre-natal mortality from the medical records where deaths in utero were coded as 'spontaneous abortions' (abortos) if they occurred before week 20 or 'stillbirths' (nacidos muertos) if after week 20. Postnatal mortality was estimated from information concerning the numbers of previous livebirths and children currently alive. Because the low-income women had higher gravidity and parity than the high-income women (gravidity=3.0 vs 2.4 pregnancies and parity=2.6 vs 2.0 livebirths respectively) and thus greater opportunity for a pre- or post-natal loss, we adjusted the mortality estimates to a common gravidity or parity. We found greater pre-natal mortality in the high than low-income group (Table 2), and this was due entirely to more deaths before week 20. Post-natal mortality, however, was markedly reduced in the high- vs low-income groups. We concluded that the Andean population was protected during prenatal life from the effects of chronic hypoxia, possibly due, in part, to factors also protecting them from altitude-associated IUGR, but that socioeconomic disadvantages outweighed these protective effects so as to raise mortality after birth.

PHYSIOLOGICAL FACTORS AFFECTING UTEROPLACENTAL O₂ DELIVERY

Normoxic pregnancy

Pregnancy affects all the determinants of O₂ delivery to the uteroplacental circulation. Ventilation rises, although this does

not normally change arterial O₂ saturation since values are already nearly maximal. Haemoglobin declines due to greater plasma than red cell mass expansion and reduces arterial O₂ content. Thus a rise in blood flow is entirely responsible for increasing O₂ delivery to the uteroplacental circulation.

The rise in uteroplacental blood flow is due, in turn, to higher cardiac output and redistribution of blood flow to favour the uteroplacental circulation. Cardiac output rises as a result of a fall in systemic vascular resistance (SVR, afterload) and a rise in blood volume (preload). The SVR fall begins in the luteal phase immediately following conception [32], most likely as the result of primary systemic vasodilation. Nitric oxide (NO) production increases [33], sympathetic tone declines, and circulating levels of the potent vasoconstrictor endothelin-1 (ET-1) fall [34]. Another key regulator is vascular endothelial growth factor (VEGF). Continual but low VEGF levels are required for endothelial cell survival [35]. But in pre-eclampsia, high levels of the membrane bound VEGF receptor 1 (sFlt-1) bind VEGF and placental like growth factor (PLGF), decrease VEGF availability, inhibit endothelial cell proliferation, and reduce vasorelaxation responses in renal arteries [36].

Redistribution of blood flow to favour the uteroplacental circulation stems primarily from profound changes which are confined to that vascular bed. Vascular resistance falls as a result of the anastomoses which develops between the ovarian branch and the main uterine artery (UA), as well as the growth and enlargement of existing vessels. Because the fall in vascular resistance in the uteroplacental bed is greater than that occurring in vessels supplied by the external iliac (EI) artery, the UA progressively 'steals' EI blood flow. Unilateral UA blood flow therefore rises from a nonpregnant value of ~10 ml/min to ~350 ml/min near term [37], a change which is greater than that experienced by any organ system following birth. In species with haemochorial placentae (e.g. most primates, rodents and guinea pigs), the vessels determining uteroplacental vascular resistance reside largely outside the uterus whereas in epitheliochorial species, the uteroplacental vessels comprise the major site of vascular resistance [38,39]. Specifically, in haemochorial species, two-thirds of the uteroplacental vascular resistance resides in the mesometrial, main UA and ovarian arteries and only one-third is located in uteroplacental channels. Since the UA makes a demonstrable contribution to uteroplacental vascular resistance in the haemochorial species under study and is the smallest vessel which can be reliably visualized in humans, our studies have focused on the UA.

UA enlargement in haemochorial species is due to alterations in its responsiveness to circulating and locally produced vasoconstrictors and vasodilators, UA distensibility and remodelling. Pregnancy raises the production of NO and other substances to augment UA vasodilator response to pharmacological agonists as well as to flow [40–42]. We and others have shown that the guinea pig UA vasoconstrictor response to alpha-adrenergic agonists is reduced [43,44]. Distensibility is also enhanced [45]. A key factor for enlarging luminal diam-

eter is UA remodelling, accompanied by compositional changes, hyperplasia and hypertrophy in all layers of the vessel wall as well [45–47]. Of note, the increase in guinea pig cellular proliferation occurs before the greatest rise in UA blood flow, suggesting that UA enlargement is a prerequisite for the flow increase to occur. The factors prompting UA growth likely involve pregnancy hormones and growth factors [48]. Since growth is more pronounced in vessels supplying the pregnant than the nonpregnant uterine horn, venous to arterial transfer of fetoplacentally derived growth factors may also be important [49]. Increased flow itself may be an important growth stimulus given that the UA's characteristic outward hypertrophic growth resembles that occurring in response to flow rather than pressure [50]. The ability of VEGF and other growth factors to stimulate NO [51] suggests that interactions between vasoactive and growth factors are likely crucial for raising UA blood flow.

Chronically hypoxic pregnancy

At high altitude, the pregnancy-associated rise in alveolar ventilation and increase in arterial O₂ saturation nearly restores arterial O₂ content to sea level values [52,53] and both relate positively to fetal weight [29,54,55]. But since birth weights are generally lower than at sea level, it is likely that reduced uteroplacental blood flow rather than diminished arterial O₂ content is chiefly responsible for the hypoxia-associated IUGR observed.

Our and others' studies show that chronic hypoxia alters systemic and uteroplacental vascular adjustments to pregnancy. In pregnant women, guinea pigs or sheep, cardiac output is lower at high vs low altitude, probably as the result of lower blood volume and/or higher SVR [56–59]. Blood volume expands normally during pregnancy at high altitude, but begins from lower non-pregnant levels such that blood volume is lower near term at high than at low altitude [58]. The higher SVR in high vs low-altitude pregnant humans or experimental animals contributes to the higher maternal blood pressures observed [16,57]. Such an increase could, in turn, be due to greater myogenic tone, altered production of local regulators of vascular tone (more vasoconstrictors and/or less vasodilators), and/or a lack of the compensatory organ remodelling that occurs in a normal pregnancy.

Factors operating both at the systemic level and within the uteroplacental circulation are likely to be important. Acute (hours) and more chronic (weeks) high-altitude exposure raise sympathetic nervous system activity and systemic catecholamine levels in nonpregnant women [60]. In addition, circulating ET-1 levels are elevated by chronic hypoxia [61]. ET-1 induced vasoconstriction appears to be an especially important contributor, given the ability of endothelin-A receptor blockade to prevent hypoxia-associated IUGR and the accompanying reduction in uteroplacental blood flow in rats [62,63].

Concerning the uteroplacental circulation, UA blood flow is one-third lower near term in pregnant high (3100 m) vs low

(1600 m) altitude residents as a result of a lesser increase in UA diameter [64]. The portion of common iliac blood flow diverted to the UA is less in pre-eclamptic women compared to normal women, suggesting even lower UA blood flows [65]. Given the importance of vessels outside the uterus in the determination of uteroplacental vascular resistance in haemochorial species, the causes of the lower UA blood flow are likely to involve the main UA, mesometrial or arcuate vessels. Of note, chronic hypoxia opposes the effects of normal pregnancy on flow vasodilation in the guinea pig UA. Thus, rather than pregnancy increasing the vasodilator response to flow, UA from chronically hypoxic animals vasoconstrict at high flow [42], resembling myometrial arteries from pre-eclamptic women in which enhanced flow vasodilation also fails to occur [66]. Chronic hypoxia also inhibits guinea pig UA growth such that there is only half as much rise in DNA synthesis as in vessels from normoxic animals [67]. Both these flow and growth alterations may stem from a lack of pregnancy-associated increase in NO. Chronic hypoxia reduces NO-dependent vasorelaxation to acetylcholine in isolated guinea pig UA rings and inhibits the pregnancy-associated increase in endothelial NOS protein (NOS III) in whole vessel homogenates [41,68]. Unknown is whether hypoxia also affects the pregnancy-associated alterations in other vasodilators (e.g. endothelial-derived hyperpolarizing factor), growth (e.g. VEGF, PIGF) or vasoactive factors (e.g. ET-1, catecholamines).

Unlike the guinea pig, a species in which hypoxia-associated IUGR occurs [57,69], a different UA response to chronic hypoxia has been observed in sheep. Sheep vary among breeds in their susceptibility to hypoxia-associated IUGR with some but not other breeds showing birth weight reductions at high altitude [56,70]. In sheep resistant to hypoxia-associated IUGR, chronic hypoxia raised the vasodilator response to acetylcholine and increase in NO production, NOS III protein and message during pregnancy to a *greater* extent in UA from high- vs low-altitude [71–73]. The converse was seen in guinea pigs as noted above, where chronic hypoxia did not alter the magnitude of acetylcholine vasodilation and reduced NO production. There are additional differences between the effects of chronic hypoxia in sheep and guinea pigs concerning the UA vasoconstrictor response to phenylephrine. Pregnancy reduced the UA vasoconstrictor sensitivity to phenylephrine similarly in chronically hypoxic vs normoxic guinea pigs. Conversely in sheep, there was a greater reduction in the chronically hypoxic vs normoxic animals as the result of decreased α 1-adrenergic receptor density, binding affinity and inositol phosphate 3 production [73–75]. Such species differences suggest that susceptibility to altitude-associated reductions in birth weight are due, at least in part, to genetic factors regulating the maternal systemic and uteroplacental circulatory responses to pregnancy [76].

Not only is there variation between species in uteroplacental vascular responses to pregnancy and hypoxia-associated IUGR, such variation also occurs within the human species. Specifically, we have shown that Tibetan women whose infants

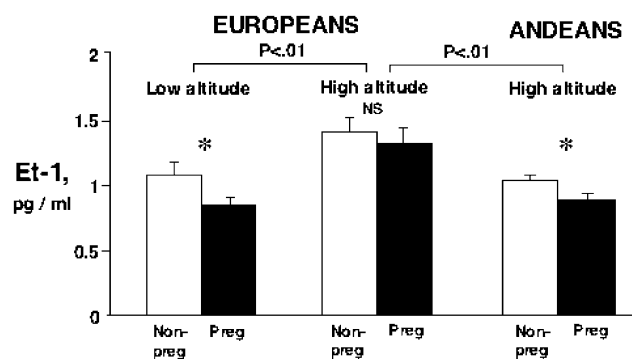


Figure 2. Andean women have lower ET-1 levels than European high-altitude (3600 m) residents when nonpregnant. Whereas values fall during pregnancy in the Andeans, there is no clear change in the European women. (*= $P < 0.05$ for comparisons of the non-pregnant vs pregnant state, and brackets= $P < 0.05$ between ancestry or altitude groups).

are protected from altitude-associated IUGR have higher UA blood flow velocity and greater lower extremity blood flow redistribution to favour the UA than Han women residing at the same elevation (3600 m) [55]. Moreover, pregnant Han women at high altitude have smaller UA diameters and lower blood flows than their low-altitude counterparts [77], like Colorado residents of high altitude [64]. Andean high-altitude pregnant women have a normal or even exaggerated fall in uteroplacental vascular resistance [78]. Preliminary data from Andean high-altitude women suggest that UA blood flow is also greater than in women of European ancestry residing at the same altitude [79]. Our preliminary data also support the hypothesis that such differences in UA blood flow may be due, in part, to differences in HIF-related genes. The potent vasoconstrictor ET-1 appears to be differentially regulated by pregnancy and chronic hypoxia in Andean vs European residents of high altitude (Figure 2). Andeans demonstrate the normal, pregnancy-associated fall in ET-1 previously reported at low altitude whereas Europeans have higher ET-1 levels and little pregnancy-associated change, like pre-eclamptic women [34]. Thus these data support the hypothesis that long-term residents of high altitude may be protected from adverse effects of chronic hypoxia on vascular responses to pregnancy via actions of HIF-targeted genes, among others.

DO GENETIC FACTORS INFLUENCE MATERNAL VASCULAR ADAPTATION TO PREGNANCY?

Population and species differences in the magnitude of hypoxia-associated IUGR suggest the involvement of genetic factors. Such genetic involvement is consistent with other studies suggesting associations between specific genetic variants and pre-eclampsia or IUGR [80–85]. In evaluating the kinds of genetic factors that might be involved, we (and others) have been struck by the observation that many of the candidate genes thus far identified are part of the HIF

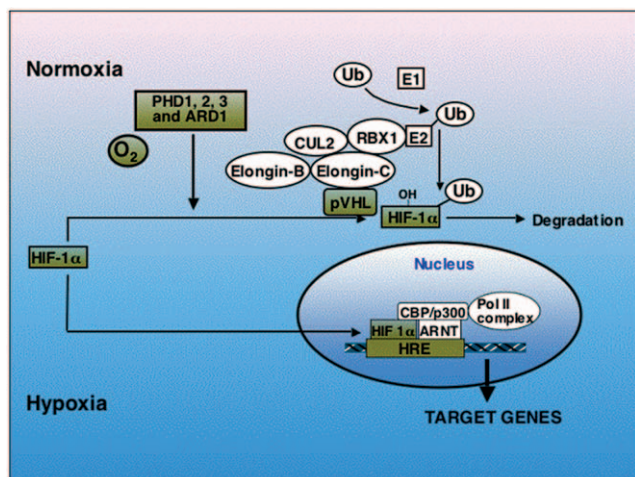


Figure 3. HIF 1- α regulation (courtesy K Stenmark).

pathway. Since this pathway regulates a majority of the hypoxia-responsive genes, we hypothesized that HIF-regulated pathways are logical targets on which selection for traits influencing variation in altitude-associated IUGR would be expected to act.

The molecular mechanisms by which HIFs influence hypoxic responses have recently been subject to extensive investigation. These studies have shown that HIF is a highly conserved heterodimer consisting of a beta subunit (the constitutive HIF1 β /ARNT complex) and one of three alpha subunits (HIF 1, 2 or 3 α). Despite continual production, degradation is sufficiently rapid that the HIF α proteins are virtually undetectable in normoxia (Figure 3). This degradation requires *trans*-4-hydroxylation at proline-564 and -402, recognition and binding by the von Hippel Lindau (VHL) protein, ubiquitination by a E3 ubiquitin ligase complex (consisting of elongin C/elongin B, cullin 2, and the RING-H2 finger protein Rbx-1), and transport to the proteasome [86]. But under hypoxia and selected other circumstances (e.g., specific oncogenes, proline hydroxylase enzyme inhibition [87], presence of large divalent metal ions or iron chelators), HIF α escapes hydroxylation and recognition by VHL. This permits HIF protein levels to rise, translocate to the nucleus, heterodimerize, and transcriptionally activate genes containing the *cis*-acting hypoxia responsive element (HRE) 5' ACGTG(C/G)3' [88].

Over 40 HIF-regulated or regulatory genes have been identified whose functions influence the vascular adjustments to hypoxia and/or pregnancy [89] (Table 3). This group comprises a number of candidate genes whose vascular effects can plausibly be linked to the vasoconstriction, endothelial damage and reduced uteroplacental blood flow characteristic of *pre-eclampsia* and IUGR [80–85]. Many (if not most) are polymorphic [i.e. exhibiting relatively common (>1 per cent) genetic variants]. Moreover, such allelic variation is likely functional insofar as it is associated with differences in levels of circulating gene products [90]. Supporting the likelihood that

Table 3. HIF genes altered by hypoxia and pregnancy

Function	Genes	Reference
HIF targets		
Vasoactive	ET-1, ECE Leptin NOS II, III Tyrosine hydroxylase 1,2; α 1-adrenergic receptor	[84,99–103] [104,105] [84,99,101,103] [106–108]
Growth	EPO, transferrin IGF (IGF2, IGFBP-3) PDGF β TGF α VEGF, Flt-1, sFlt, KDR, neuropilin 1-2, PIGF	[109–111] [112–115] [116,117] [8,118] [36,102,119–126]
Inflammation	Interleukins 1, 6; TNF α	[127–132]
HIF regulatory	HIF1-3 α , HIF1- α , ARNT2-3 ARD1 Cul2, JAB1/CSN5, RBX1 PHD1, 2, 3 VHL	[8,133–135] [136] [137–139] [140,141] [7,142]

Notes: NOSIII is hypoxia but not HIF-regulated. Cul2 and RBX1 are constitutive. Effects of pregnancy on Cul2, RBX1, and PHD are unknown.

such genes are expressed in the uteroplacental vascular bed, HIF or HIF-regulatory gene expression is altered in placentae from pregnancies complicated by *pre-eclampsia* and/or IUGR compared with normal pregnancy [8,9].

GENOMIC APPROACHES FOR IDENTIFYING CANDIDATE GENES

Genomic approaches have been infrequently applied to pregnancy complications but offer considerable power for determining interindividual variation in risk factors for complex diseases [91,92]. Such approaches include ones which take advantage of the possibility that natural selection has acted to differentiate one population, in this case Andeans, from other groups with shorter duration of high-altitude exposure with respect to genes influencing uteroplacental blood flow and fetal growth. The principle underlying these approaches stems from the observation that since geographic separation of human populations is relatively recent,¹ most genetic variation is shared among all groups with only ~15 per cent differing between major continental regions [93]. Thus, comparatively small differences in allele frequency are expected across most genes but not, importantly, those that have been acted upon by natural selection [94]. For example, the Duffy null allele

¹ 100–60 000 years ago (kya) for African/non-African, ~50 kya for Asian/European, ~20–15 kya for Asian/American separation.

Table 4. Allele frequencies, differentials, and LSBLs for 10k-WGA SNPs near HIF-regulated/regulatory genes

Gene	SNP no.	Allele frequencies			Quechua–Nahua differential	Quechua LSBL
		Quechua	Nahua	Asian		
NOSII	1	0.425	0.588	0.500	0.163	0.075
NOSII	2	0.425	0.650	0.525	0.225 ^b	0.100
α 1-adrenergic receptor		0.100	0.250	0.350	0.150	0.150 ^b
endothelin		0.800	0.525	0.175	0.275 ^a	0.275 ^a
FLT1	1	0.825	0.900	0.600	0.075	0.000
TGF α		0.225	0.250	0.450	0.025	0.025
cullin 2		0.175	0.325	0.200	0.150	0.025
neuropilin 2	1	0.400	0.421	0.600	0.021	0.021
neuropilin 2	2	0.050	0.053	0.150	0.003	0.003
neuropilin 1		0.500	0.421	0.368	0.079	0.079
PHD3	1	0.775	0.625	0.500	0.150	0.150 ^b
PHD3	2	0.361	0.214	0.250	0.147	0.111
PHD3	3	0.425	0.400	0.075	0.025	0.025

^a $P < 0.05$.

^b $0.05 < P < 0.10$.

(FY^*0) which provides protection from the *Plasmodium vivax* form of malaria has reached very high frequencies across sub-Saharan Africa in the last several thousand years, while not being present outside Africa [95]. Divergence for FY^*0 and markers near the functional site is detected as high F_{ST} levels (i.e., the proportion of the total genetic variation that is due to differences among groups) [96]. We recently extended the F_{ST} approach to quantify locus-specific divergence using a measure termed locus-specific branch length (LSBL) [97]. While the F_{ST} approach evaluates if any one or more of the populations under consideration have undergone dramatic changes in allele frequency, the LSBL approach provides the ability to geometrically isolate the population in which the allele frequency change occurred. These F_{ST} and LSBL based methods provide exciting breakthroughs for identifying candidate genes in pathways thought affected by recent directional or balancing selection [92,94,98].

As a preliminary test as to whether any of the candidate genes listed in Table 3 are implicated in the population differences observed in altitude-associated IUGR, we generated an empirical distribution for the levels of LSBL in Andean high-altitude residents and, for comparative purposes, indigenous American groups not residing at high altitude. In addition, we included East Asians as a group which is likely to share a recent (relative to other human populations) common ancestor [97]. We used a new genotyping method developed by Affymetrix (10K-WGA mapping array, Santa Clara, CA, USA) that permits whole genome amplification and generation of allele frequencies for 11 555 single nucleotide polymorphisms (SNPs) located throughout the genome. Using the Affymetrix 10K-WGA chip, we examined several populations, including East Asians and two indigenous Central and South American groups (Nahua and Andean), both of which have previously been shown to have very low (<1 per cent) African or European admixture. These empirical distributions of F_{ST} and LSBL can then be used to quantify how different any

particular gene is from the range of values typically seen for the population comparisons being made.

For a preliminary evaluation of branch lengths for the candidate genes listed in Table 3, we screened for genes located within 40 kb of the 11 555 SNPs inventoried on the 10K-WGA chip. Nine such genes were identified (Table 4). Shown are the allele frequencies for the SNPs near each of these nine genes, the difference in allele frequency, and the LSBL for Andeans (Quechua from Peru) and two related populations [Nahua from Mexico and East Asians (Chinese and Japanese living in the US)]. Of these nine, remarkably, nearly half (44 per cent) had one SNP which was in the highest 90th percentile of the distribution of all SNPs on the 10K-WGA. These genes were NOSII, the alpha1-adrenergic receptor, endothelin, and PHD3, all of which are important candidate genes. To phrase this differently, there is a less than 10 per cent chance that genetic variation near these four genes was within the range exhibited in the low-altitude control populations (the Nahua and East Asians) and a <5 per cent chance that variation near the ET-1 gene was within the expected range.

DIRECTIONS FOR FUTURE STUDY

Thus, the available data suggest that UA blood flow is lower during pregnancy at high altitude in settings where IUGR is most pronounced. Further the variation in hypoxia-associated IUGR that is evident both within as well as between species suggests important avenues for future studies designed to address the contribution of genetic factors to maternal vascular adaptation to pregnancy. Future studies are required for assaying additional SNPs near the candidate, HIF-targeted genes to verify the existence of recent natural selection in Andean populations and for testing functionality. Such studies represent a novel and, as yet, relatively unexplored approach

for achieving an integrated understanding of the physiological and genetic bases for pregnancy complications. Such an approach can also be expected to yield new predictive tests as

well as therapies for treatment designed to alleviate suffering at this most vulnerable period of life.

ACKNOWLEDGEMENTS

Our appreciation is extended to the many subjects who have participated in our studies of pregnancy at high altitudes in Colorado, Peru, Tibet and Bolivia and to the technical staff at the Women's Health and Colorado High-Altitude Research Centers, the Bolivian High-Altitude Biology Institute, the Pennsylvania Department of Anthropology and the other sites which have cooperated with us in the past. We especially thank Ms Wendy MacCannell for her help with the preparation of this article and Rhonda Mouser for the conduct of the endothelin-1 assays. Grant support was provided by NIH TW01188, HL60131, HL14985.

REFERENCES

- [1] Niermeyer S, Zamudio S, Moore LG. The people. In: Hornbein T, Schoene RB, editors. Adaptations to hypoxia. New York (NY): Marcel Dekker and Co; 2001, p. 43–100.
- [2] Lichty JL, Ting R, Bruns PD, Dyar E. Studies of babies born at high altitude. *Am J Dis Child* 1957;93:666–9.
- [3] Moore LG. Small babies and big mountains: John Lichty solves a Colorado mystery in Leadville. In: Reeves JT, Grover FT, editors. Attitudes on altitude. Boulder: Univ Colorado Press; 2001b, p. 137–59.
- [4] Lubchenco LO. Intrauterine growth as estimated from liveborn birth-weight data at 24 to 42 weeks of gestation. *Pediatrics* 1963;32:793–800.
- [5] Moore LG, Hershey DW, Jahnigen D, Bowes W Jr. The incidence of pregnancy-induced hypertension is increased among Colorado residents at high altitude. *Am J Obstet Gynecol* 1982a;144:423–9.
- [6] Ahmed A, Dunk C, Ahmad S, Khaliq A. Regulation of placental vascular endothelial growth factor (VEGF) and placenta growth factor (PLGF) and soluble Flt-1 by oxygen—a review. *Placenta* 2000;21:S16–24.
- [7] Genbacev O, Krtolica A, Kaelin W, Fisher SJ. Human cytotrophoblast expression of the von Hippel-Lindau protein is downregulated during uterine invasion in situ and upregulated by hypoxia in vitro. *Dev Biol* 2001;233:526–36.
- [8] Caniggia I, Winter JL. Hypoxia inducible factor-1: oxygen regulation of trophoblast differentiation in normal and pre-eclamptic pregnancies—a review. *Placenta* 2002;23:S47–57.
- [9] Rajakumar A, Doty K, Daftary A, Harger G, Conrad KP. Impaired oxygen-dependent reduction of HIF-1 α and -2 α proteins in pre-eclamptic placentae. *Placenta* 2003;24:199–208.
- [10] Kaufmann P, Black S, Huppertz B. Endovascular trophoblast invasion: implications for the pathogenesis of intrauterine growth retardation and preeclampsia. *Biol Reprod* 2003;69:1–7.
- [11] Jensen GM, Moore LG. The effect of high altitude and other risk factors on birthweight: independent or interactive effects?. *Am J Public Health* 1997;87:1003–7.
- [12] Giussani DA, Seamus P, Anstee S, Barker DJP. Effects of altitude versus economic status on birth weight and body shape at birth. *Pediatr Res* 2001;49:490–4.
- [13] Mortola JP, Frappell PB, Aguero L, Armstrong K. Birth weight and altitude: a study in Peruvian communities. *J Pediatr* 2000;136:324–9.
- [14] Unger C, Weiser JK, McCullough RE, Keefer S, Moore LG. Altitude, low birth weight, and infant mortality in Colorado. *J Am Med Assoc* 1988;259:3427–32.
- [15] Krampfl E, Lees C, Bland JM, Dorado JE, Gonzalo M, Campbell S. Fetal biometry at 4300 m compared to sea level in Peru. *Ultrasound Obstet Gynecol* 2000;16:9–18.
- [16] Palmer SK, Moore LG, Young DA, Cregger B, Berman JC, Zamudio S. Altered blood pressure course during normal pregnancy and increased preeclampsia at high altitude (3100 meters) in Colorado. *Am J Obstet Gynecol* 1999;180:1161–8.
- [17] Keyes LE, Armaza JF, Niermeyer S, Vargas E, Young D, Villena M et al. Intrauterine growth restriction, preeclampsia and intrauterine mortality at high altitude in Bolivia. *Pediatr Res* 2003 in press.
- [18] Niermeyer S. Cardiopulmonary transition in the high altitude neonate. *High Alt Med Biol* 2003;4:225–39.
- [19] McCullough RE, Reeves JT, Liljegren RL. Fetal growth retardation and increased infant mortality at high altitude. *Arch Environ Health* 1977;32:36–40.
- [20] PAHO. Health conditions in the Americas. Scientific publication No. 549. Washington (DC, USA): World Health Organization (Pan American Health Organization), 1994.
- [21] Wiley AS. Neonatal size and infant mortality at high altitude in the western Himalaya. *Am J Phys Anthropol* 1994;94:289–305.
- [22] Bolivia Ro. Encuesta Nacional de Demografía y Salud. Calverton (MD, USA): Macro International/DHS+ Program, 1998.
- [23] Giussani D. High altitude and rural living are associated with increased infant mortality in Bolivia. *J Soc Gynecol Invest* 2002;9:292A.
- [24] PAHO. *Epidemiol Bull* 1999;20(3):14–9.
- [25] Haas JD. Human adaptability approach to nutritional assessment: a Bolivian example. *Fed Proc* 1981;40:2577–82.
- [26] Moore LG. Human genetic adaptation to high altitude. *High Alt Med Biol* 2001a;2:257–79.
- [27] Moore LG, Young DY, McCullough RE, Droma TS, Zamudio S. Tibetan protection from intrauterine growth restriction (IUGR) and reproductive loss at high altitude. *Am J Human Biol* 2001a;13:635–44.
- [28] Weinstein RS, Haas JD. Early stress and later reproductive performance under conditions of malnutrition and high altitude hypoxia. *Med Anthropol* 1977;1:25–54.
- [29] Moore LG, Rounds SS, Jahnigen D, Grover RF, Reeves JT. Infant birth weight is related to maternal arterial oxygenation at high altitude. *J Appl Physiol* 1982b;52:695–9.
- [30] Chakraborty R, Barton SA, Ferrell RE, Schull WJ. Ethnicity determination by names among the Aymara of Chile and Bolivia. *Hum Biol* 1989;61:159–77.
- [31] Moore LG, Armaza F, Keyes L, Borth R, Niermeyer S, Villena M et al. Andean compared with European babies are protected from altitude-associated intrauterine growth restriction (IUGR). *High Alt Med Biol* 2002;4:460 (abstract).
- [32] Chapman AB, Abraham WT, Osorio FV, Merouani A, Zamudio S, Young DA et al. Temporal relationships between hormonal and hemodynamic changes in early human pregnancy. *Kidney Int* 1998;54:2056–63.
- [33] Cadnapaphornchai MA, Ohara M, Morris KG Jr, Knotek M, Rogachev B, Ladtkow T et al. Chronic NOS inhibition reverses systemic vasodilation and glomerular hyperfiltration in pregnancy. *Am J Physiol Renal Physiol* 2001;280:F592–8.
- [34] Furuhashi N, Kimura H, Nagae H, Yajima A. Maternal plasma endothelin levels and fetal status in normal and preeclamptic pregnancies. *Gynecol Obstet Invest* 1995;39:88–92.
- [35] Luttun A, Carmeliet P. Soluble VEGF receptor Flt1: the elusive preeclampsia factor discovered?. *J Clin Invest* 2003;111:600–2.
- [36] Maynard SE, Min J-Y, Merchan J, Lim K-H, Li J, Mondal S et al. Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *J Clin Invest* 2003;111:649–58.
- [37] Palmer SK, Zamudio S, Coffin C, Parker S, Stamm E, Moore LG. Quantitative estimation of human uterine artery blood flow and pelvic blood flow redistribution in pregnancy. *Obstet Gynecol* 1992;80:1000–6.
- [38] Moll W, Kunzel W. The blood pressure in arteries entering the placenta of guinea pigs, rats, rabbits, and sheep. *Pflügers Arch* 1973;338:125–31.
- [39] Stock M, Metcalfe J. Maternal physiology during gestation. In: Knobil E, Neil J, editors. The physiology of reproduction. New York: Raven Press; 1994, p. 947–83.
- [40] Weiner CP, Thompson LP, Liu KZ, Herrig JE. Endothelium-derived relaxing factor and indomethacin-sensitive contracting factor alter arterial contractile responses to thromboxane during pregnancy. *Am J Obstet Gynecol* 1992;166:1171–8.
- [41] White MM, McCullough RE, Dyckes R, Robertson AD, Moore LG. Chronic hypoxia, pregnancy, and endothelium-mediated relaxation in guinea pig uterine and thoracic arteries. *Am J Physiol Heart Circ Physiol* 2000;278:H2069–75.

- [42] Mateev S, Sillau AH, Mouser R, McCullough RE, White MM, Young DA et al. Chronic hypoxia opposes pregnancy-induced increase in uterine artery vasodilator response to flow. *Am J Physiol Heart Circ Physiol* 2003;284:H820-9.
- [43] Weiner C, Liu KZ, Thompson L, Herrig J, Chestnut D. Effect of pregnancy on endothelium and smooth muscle: their role in reduced adrenergic sensitivity. *Am J Physiol* 1991;261:H1275-83.
- [44] White MM, McCullough RE, Dyckes R, Robertson AD, Moore LG. Effects of pregnancy and chronic hypoxia on contractile responsiveness to α -1-adrenergic stimulation. *J Appl Physiol* 1998;85:2322-9.
- [45] Mateev S, Mouser R, Min L, Moore LG. Chronic hypoxia augments uterine artery distensibility and increases myogenic tone during pregnancy. *J Soc Gynecol Investig* 2002;9:299A.
- [46] Cipolla M, Osol G. Hypertrophic and hyperplastic effects of pregnancy on the rat uterine arterial wall. *Am J Obstet Gynecol* 1994;171:805-11.
- [47] Keyes LE, Majack R, Dempsey EC, Moore LG. Pregnancy stimulation of DNA synthesis and uterine blood flow in the guinea pig. *Pediatr Res* 1997;41:708-15.
- [48] Keyes LE, Moore LG, Walchak SJ, Dempsey EC. Pregnancy-stimulated growth of vascular smooth muscle cells: importance of protein kinase C-dependent synergy between estrogen and platelet-derived growth factor. *J Cell Physiol* 1996;166:22-32.
- [49] Celia G, Osol G. VEGF modulates uterine veno-arterial communication through a PLC-PKC signaling cascade that is independent of nitric oxide. *J Soc Gynecol Investig* 2002;9:86A.
- [50] Mulvany MJ, Baumbach GL, Aalkjaer C, Heagerty AM, Korsgaard N, Schiffrin EL et al. Vascular remodeling. *Hypertension* 1996;28:505-6.
- [51] Ni Y, May V, Braas K, Osol G. Pregnancy augments uteroplacental vascular endothelial growth factor gene expression and vasodilator effects. *Am J Physiol Heart Circ Physiol* 1997;273:H938-44.
- [52] Moore LG, Jahnigen D, Rounds SS, Reeves JT, Grover RF. Maternal hyperventilation helps preserve arterial oxygenation during high-altitude pregnancy. *J Appl Physiol* 1982;52:690-4.
- [53] McAuliffe F, Kametas N, Krampf E, Ernsting J, Nicolaides K. Blood gases in pregnancy at sea level and at high altitude. *Br J Obstet Gynaecol* 2001;108:980-5.
- [54] Moore LG, Brodeur P, Chumbe O, D'Brot J, Hofmeister S, Monge C. Maternal hypoxic ventilatory response, ventilation, and infant birth weight at 4,300 m. *J Appl Physiol* 1986;60:1401-6.
- [55] Moore LG, Zamudio S, Zhuang J, Sun S, Droma T. Oxygen transport in Tibetan women during pregnancy at 3658 m. *Am J Phys Anthropol* 2001b;114:42-53.
- [56] Kitanaka T, Gilbert RD, Longo LD. Maternal responses to long-term hypoxemia in sheep. *Am J Physiol* 1989;256:R1340-7.
- [57] Harrison GL, Moore LG. Systemic vascular reactivity during high-altitude pregnancy. *J Appl Physiol* 1990;69:201-6.
- [58] Zamudio S, Palmer SK, Dahms TE, Berman JC, McCullough RG, McCullough RE et al. Blood volume expansion, preeclampsia, and infant birth weight at high altitude. *J Appl Physiol* 1993;75:1566-73.
- [59] Kametas NA, Savvidou MD, Donald AE, McAuliffe F, Nicolaides KH. Flow-mediated dilatation of the brachial artery in pregnancy at high altitude. *Br J Obstet Gynaecol* 2002;109:930-7.
- [60] Mazzeo RS, Child A, Butterfield GE, Mawson JT, Zamudio S, Moore LG. Catecholamine response during 12 days of high-altitude exposure (4,300 m) in women. *J Appl Physiol* 1998;84:1151-7.
- [61] Morganti A, Giussani M, Sala C, Gazzano G, Marana I, Pierini A et al. Effects of exposure to high altitude on plasma endothelin-1 levels in normal subjects. *J Hypertens* 1995;13:859-65.
- [62] Thaete LG, Neerhof MG, Caplan MS. Endothelin receptor A antagonism prevents hypoxia-induced intrauterine growth restriction in the rat. *Am J Obstet Gynecol* 1997;176:73-6.
- [63] Thaete LG, Neerhof MG. Contribution of endothelin to the regulation of uterine and placental blood flow during nitric oxide synthase inhibition in the pregnant rat. 7th International Conference on Endothelin, 2001, p. 222.
- [64] Zamudio S, Palmer SK, Droma T, Stamm E, Coffin C, Moore LG. Effect of altitude on uterine artery blood flow during normal pregnancy. *J Appl Physiol* 1995a;79:7-14.
- [65] Zamudio S, Palmer SK, Dahms TE, Berman JC, Young DA, Moore LG. Alterations in uteroplacental blood flow precede hypertension in preeclampsia at high altitude. *J Appl Physiol* 1995b;79:15-22.
- [66] Kublickiene KR, Lindblom B, Kruger K, Nisell H. Preeclampsia: evidence for impaired shear stress-mediated nitric oxide release in uterine circulation. *Am J Obstet Gynecol* 2000;183:160-6.
- [67] Rockwell LC, Keyes LE, Moore LG. Chronic hypoxia diminishes pregnancy-associated DNA synthesis in guinea pig uteroplacental arteries. *Placenta* 2000;21:313-9.
- [68] White MM, Moore LG, Mouser R, Le Cras TD. Effect of pregnancy and chronic hypoxia on endothelial nitric oxide synthase (NOSIII) protein expression in guinea pig uterine and thoracic arteries. *J Soc Gynecol Investig* 2001;8:187A.
- [69] Gilbert RD, Cummings LA, Juchau MR, Longo LD. Placental diffusing capacity and fetal development in exercising or hypoxic guinea pigs. *J Appl Physiol* 1979;46:828-34.
- [70] Jacobs R, Roninson JS, Owens JA, Falconer J, Webster ME. The effect of prolonged hypobaric hypoxia on growth of fetal sheep. *J Dev Physiol* 1988;10:97-112.
- [71] Zhang L, Xiao D, Bouslough DB. Long-term high-altitude hypoxia increases plasma nitrate levels in pregnant ewes and their fetuses. *Am J Obstet Gynecol* 1998;179:1594-8.
- [72] Xiao D, Liu Y, Pearce WJ, Zhang L. Endothelial nitric oxide release in isolated perfused ovine uterine arteries: effect of pregnancy. *Eur J Pharmacol* 1999;367:223-30.
- [73] Xiao D, Pearce WJ, Zhang L. Pregnancy enhances endothelium-dependent relaxation of ovine uterine artery: role of NO and intracellular Ca(2+). *Am J Physiol Heart Circ Physiol* 2001;281:H183-90.
- [74] Hu XQ, Longo LD, Gilbert RD, Zhang L. Effects of long-term high-altitude hypoxemia on alpha 1-adrenergic receptors in the ovine uterine artery. *Am J Physiol* 1996;270:H1001-7.
- [75] Hu XQ, Zhang L. Chronic hypoxia suppresses pharmacomechanical coupling of the uterine artery in near-term pregnant sheep. *J Physiol (Lond)* 1997;499:551-9.
- [76] White MM, Zhang L. Effects of chronic hypoxia on maternal vascular changes in guinea pig and ovine pregnancy. *High Alt Med Biol* 2003;4:157-69.
- [77] Chen D, Zhou X, Zhu Y, Zhu T, Wang J. Comparison study on uterine and umbilical artery blood flow during pregnancy at high altitude and at low altitude. *Zhonghua Fu Chan Ke Za Zhi* 2002;37:69-71.
- [78] Krampf E, Espinoza-Dorado J, Lees CC, Moscoso G, Bland JM. Maternal uterine artery Doppler studies at high altitude and sea level. *Ultrasound Obstet Gynecol* 2001;18:578-82.
- [79] Rockwell LC, Moore LG. Maternal physiological adaptation to pregnancy. *Am J Human Biol* 2003 in press.
- [80] Ward K, Hata A, Jeunemaitre X, Helin C, Nelson L, Namikawa C et al. A molecular variant of angiotensinogen associated with preeclampsia [see comments]. *Nat Genet* 1993;4:59-61.
- [81] Angrimsson R, Hayward C, Nadaud S, Baldursdottir A, Walker JJ, Liston WA et al. Evidence for a familial pregnancy-induced hypertension locus in the eNOS-gene region. *Am J Hum Genet* 1997;61:354-62.
- [82] Lie RT, Rasmussen S, Brunborg H, Gjessing HK, Lie-Nielsen E, Irgens LM. Fetal and maternal contributions to risk of pre-eclampsia: population based study. *BMJ* 1998;316:1343-7.
- [83] Hubel CA, Roberts JM, Ferrell RE. Association of pre-eclampsia with common coding sequence variations in the lipoprotein lipase gene. *Clin Genet* 1999;56:289-96.
- [84] Savvidou MD, Valance PJT, Nicolaides KH, Hingorani AD. Endothelial nitric oxide synthase gene polymorphism and maternal vascular adaptation to pregnancy. *Hypertension* 2001;38:1289-93.
- [85] Lindsay RS, Kobes S, Knowler WC, Hanson RL. Genome-wide linkage analysis assessing parent-of-origin effects in the inheritance of birth weight. *Hum Genet* 2002;110:503-9.
- [86] Safran M, Kaelin WG Jr. HIF hydroxylation and the mammalian oxygen-sensing pathway. *J Clin Invest* 2003;111:779-83.
- [87] Huang J, Zhao Q, Mooney SM, Lee FS. Sequence determinants in hypoxia-inducible factor-1alpha for hydroxylation by the prolyl hydroxylases PHD1, PHD2, and PHD3. *J Biol Chem* 2002;277:39792-800.
- [88] Semenza GL. Regulation of mammalian O₂ homeostasis by hypoxia-inducible factor 1. *Annu Rev Cell Dev Biol* 1999;15:551-78.
- [89] Ratcliffe PJ, O'Rourke JF, Maxwell PH, Pugh CW. Oxygen sensing, hypoxia-inducible factor-1 and the regulation of mammalian gene expression. *J Exp Biol* 1998;201:1153-62.
- [90] Renner W, Kotschan S, Hoffmann C, Obermayer-Pietsch B, Pilger E. A common 936 C/T mutation in the gene for vascular endothelial growth factor is associated with vascular endothelial growth factor plasma levels. *J Vasc Res* 2000;37:443-8.
- [91] Black WC, Baer CF, Antolin MF, DuTeau NM. Population genomics: genome-wide sampling of insect populations. *Annu Rev Entomol* 2001;46:441-69.

- [92] Schlotterer C. Hitchhiking mapping—functional genomics from the population genetics perspective. *Trends Genet* 2003;19:32–8.
- [93] Lewontin RC. Ken-Ichi Kojima September 17, 1930–November 14, 1971. *Genetics* 1972;71(Suppl 2):s89–90.
- [94] Akey JM, Zhang G, Zhang K, Jin L, Shriver MD. Interrogating a high-density SNP map for signatures of natural selection. *Genome Res* 2002;12:1805–14.
- [95] Cavali-Sforza LL. Population structure and human evolution. *Proc R Soc Lond B Biol Sci* 1966;164:362–79.
- [96] Hamblin MT, Thompson EE, Di Rienzo A. Complex signatures of natural selection at the Duffy blood group locus. *Am J Hum Genet* 2002;70:369–83.
- [97] Shriver MD, Kennedy GC, Parra EJ, Lawson HA, Huang J, Makova K et al. The genomic distribution of human population substructure. 2003; in review.
- [98] Bamshad M, Wooding SP. Signatures of natural selection in the human genome. *Nat Rev Genet* 2003;4:99–111.
- [99] McQuillan LP, Leung GK, Marsden PA, Kostyk SK, Kourembanas S. Hypoxia inhibits expression of eNOS via transcriptional and posttranslational mechanisms. *Am J Physiol* 1994;267:H1921–7.
- [100] Bodi I, Bishopric NH, Discher DJ, Wu X, Webster KA. Cell-specificity and signaling pathway of endothelin-1 gene regulation by hypoxia. *Cardiovasc Res* 1995;30:975–84.
- [101] Melillo G, Musso T, Sica A, Taylor LS, Cox GW, Varesio L. A hypoxia-responsive element mediates a novel pathway of activation of the inducible nitric oxide synthase promoter. *J Exp Med* 1995;182:1683–93.
- [102] Camenisch G, Stroka DM, Gassmann M, Wenger RH. Attenuation of HIF-1 DNA-binding activity limits hypoxia-inducible endothelin-1 expression. *Pflügers Arch Gesamte Physiol Menschen Tiere* 2001;443:240–9.
- [103] Kimura H, Esumi H. Reciprocal regulation between nitric oxide and vascular endothelial growth factor in angiogenesis. *Acta Biochim Pol* 2003;50:49–59.
- [104] Grosfeld A, Andre J, Hauguel-De Mouzon S, Berra E, Pouyssegur J, Guerre-Millo M. Hypoxia-inducible factor 1 transactivates the human leptin gene promoter. *J Biol Chem* 2002;277:42953–7.
- [105] Meissner U, Ostreicher I, Allabauer I, Rascher W, Dotsch J. Synergistic effects of hypoxia and insulin are regulated by different transcriptional elements of the human leptin promoter. *Biochem Biophys Res Commun* 2003;303:707–12.
- [106] Czyzyk-Krzeska MF, Bayliss DA, Lawson EE, Millhorn DE. Regulation of tyrosine hydroxylase gene expression in the rat carotid body by hypoxia. *J Neurochem* 1992;58:1538–46.
- [107] Schobel HP, Fischer T, Heuszer K, Geiger H, Schmieder RE. Preeclampsia—a state of sympathetic overactivity. *N Engl J Med* 1996;335:1480–5.
- [108] Smiley RM, Finster M. Do receptors get pregnant too? Adrenergic receptor alterations in human pregnancy. *J Matern Fetal Med* 1996;5:106–14.
- [109] Goldberg MA, Gaut CC, Bunn HF. Erythropoietin mRNA levels are governed by both the rate of gene transcription and posttranscriptional events. *Blood* 1991;77:271–7.
- [110] Rolfes A, Kvietikova I, Gassmann M, Wenger RH. Oxygen-regulated transferrin expression is mediated by hypoxia-inducible factor-1. *J Biol Chem* 1997;272:20055–62.
- [111] Ostlund E, Lindholm H, Hensen A, Fried G. Fetal erythropoietin and endothelin-1: relation to hypoxia and intrauterine growth retardation. *Acta Obstet Gynecol Scand* 2000;79:276–82.
- [112] Zelzer E, Levy Y, Kahana C, Shilo BZ, Rubinstein M, Cohen B. Insulin induces transcription of target genes through the hypoxia-inducible factor HIF-1 α /ARNT. *EMBO J* 1998;17:5085–94.
- [113] Zamudio S, Broad E, Niccoli S, Bhatia S, Tissot Van Patot MC, Faessen G et al. IGFBP-1 and IGF-1 in high vs. moderate altitude pregnancy. *J Soc Gynecol Invest* 2000;7:252A.
- [114] Constancia M, Hemberger M, Hughes JM, Dean W, Ferguson-Smith A, Fundele R et al. Placental-specific IGF-II is a major modulator of placental and fetal growth. *Nature* 2002;417:945–8.
- [115] Martinez-Chequer JC, Stouffer RL, Hazzard TM, Patton PE, Molskness TA. Insulin-like growth factors-1 and -2, but not hypoxia, synergize with gonadotropin hormone to promote vascular endothelial growth factor—a secretion by monkey granulosa cells from preovulatory follicles. *Biol Reprod* 2003;68:1112–8.
- [116] Heinig J, Wilhelm S, Muller H, Briese V, Bittorf T, Brock J. Determination of cytokine mRNA-expression in term human placenta of patients with gestational hypertension, intrauterine growth retardation and gestational diabetes mellitus using polymerase chain reaction. *Zentralbl Gynakol* 2000;122:413–8.
- [117] Gorlach A, Diebold I, Schini-Kerth VB, Brechner-Pfannschmidt U, Roth U, Brandes RP et al. Thrombin activates the hypoxia-inducible factor-1 signaling pathway in vascular smooth muscle cells: Role of the p22(phox)-containing NADPH oxidase. *Circ Res* 2001;89:47–54.
- [118] Shweiki D, Itin A, Soffer D, Keshet E. Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. *Nature* 1992;359:843–5.
- [119] Gleadle JM, Ebert BL, Firth JD, Ratcliffe PJ. Regulation of angiogenic growth factor expression by hypoxia, transition metals and chelating agents. *Am J Physiol* 1995;268:C1362–8.
- [120] Gerber HP, Condorelli F, Park J, Ferrara N. Differential transcriptional regulation of the two vascular endothelial growth factor receptor genes. Flt-1, but not Flk-1/KDR, is up-regulated by hypoxia. *J Biol Chem* 1997;272:23659–67.
- [121] Khaliq A, Dunk C, Jiang J, Shams M, Li XF, Acevedo C et al. Hypoxia down-regulates placenta growth factor, whereas fetal growth restriction up-regulates placenta growth factor expression: molecular evidence for “placental hyperoxia” in intrauterine growth restriction. *Lab Invest* 1999;79:151–70.
- [122] Halder JB, Zhao X, Soker S, Paria BC, Klagsbrun M, Das SK et al. Differential expression of VEGF isoforms and VEGF(164)-specific receptor neuropilin-1 in the mouse uterus suggests a role for VEGF(164) in vascular permeability and angiogenesis during implantation. *Genesis* 2000;26:213–24.
- [123] Neufeld G, Kessler O, Herzog Y. The interaction of Neuropilin-1 and Neuropilin-2 with tyrosine-kinase receptors for VEGF. *Adv Exp Med Biol* 2000;515:81–90.
- [124] Ong CY, Liao AW, Cacho AM, Spencer K, Nicolaidis KH. First-trimester maternal serum levels of placenta growth factor as predictor of preeclampsia and fetal growth restriction. *Obstet Gynecol* 2001;98:608–11.
- [125] Elvert G, Kappel A, Heidenreich R, Englemeier U, Lanz S, Acker T et al. Cooperative interaction of hypoxia-inducible factor-2 α (HIF-2 α) and Ets-1 in the transcriptional activation of vascular endothelial growth factor receptor-2 (Flk-1). *J Biol Chem* 2003;278:7520–30.
- [126] Trollmann R, Amann K, Schoof E, Beinder E, Wenzel D, Rascher W et al. Hypoxia activates the human placental vascular endothelial growth factor system in vitro and in vivo: up-regulation of vascular endothelial growth factor in clinically relevant hypoxic ischemia in birth asphyxia. *Am J Obstet Gynecol* 2003;188:517–23.
- [127] Benyo DF, Miles TM, Conrad KP. Hypoxia stimulates cytokine production by villous explants from the human placenta. *J Clin Endocrinol Metab* 1997;82:1582–8.
- [128] Hellwig-Bürgel T, Rutkowski K, Metzgen E, Fandrey J, Jelkmann W. Interleukin-1 β and tumor necrosis factor- α stimulate DNA binding of hypoxia-inducible factor-1. *Blood* 1999;94:1561–7.
- [129] Lea RG, Riley SC, Antipatis C, Hannah L, Ashworth CJ, Clark DA et al. Cytokines and the regulation of apoptosis in reproductive tissues: a review. *Am J Reprod Immunol* 1999;42:100–9.
- [130] Rinehart BK, Terrone DA, Lagoo-Deenadayalan S, Barber WH, Hale EA, Martin JN et al. Expression of the placental cytokines tumor necrosis factor alpha, interleukin 1beta, and interleukin 10 is increased in preeclampsia. *Am J Obstet Gynecol* 1999;181:915–20.
- [131] Thornton RD, Lane P, Borghaei RC, Pease EA, Caro J, Mochan E. Interleukin 1 induces hypoxia-inducible factor 1 in human gingival and synovial fibroblasts. *Biochem J* 2000;350:307–12.
- [132] Coussons-Read ME, Mazzeo RS, Whitford MH, Schmitt M, Moore LG, Zamudio S. High altitude residence during pregnancy alters cytokine and catecholamine levels. *Am J Reprod Immunol* 2002;48:344–54.
- [133] Chan WK, Yao G, Gu YZ, Bradfield CA. Cross-talk between the aryl hydrocarbon receptor and hypoxia inducible factor signaling pathways. *J Biol Chem* 1999;274:12115–23.
- [134] Daikoku T, Matsumoto H, Gupta RA, Das SK, Gassmann M, DuBois RN et al. Expression of hypoxia-inducible factors in the peri-implantation mouse uterus is regulated in a cell-specific and ovarian steroid hormone-dependent manner. Evidence for differential function of HIFs during early pregnancy. *J Biol Chem* 2003;278:7683–91.

- [135] Rajakumar A. The hypoxia inducible transcription factor, HIF-1 α , overexpressed in preeclamptic placentas is capable of binding to the hypoxia response element. *J Soc Gynecol Investig* 2003;10:304A.
- [136] Jeong JW, Bae MK, Ahn MY, Kim SH, Sohn TK, Bae MH et al. Regulation and destabilization of HIF-1 α by ARD1-mediated acetylation. *Cell* 2002;111:709–20.
- [137] Bae MK, Ahn MY, Jeong JW, Bae MH, Lee YM, Bae SK et al. Jab1 interacts directly with HIF-1 α and regulates its stability. *J Biol Chem* 2002;277:9–12.
- [138] Cope GA, Suh GS, Aravind L, Schwarz SE, Zipursky SL, Koonin EV et al. Role of predicted metalloprotease motif of Jab1/Csn5 in cleavage of Nedd8 from Cull1. *Science* 2002;298:608–11.
- [139] Ietta F, Todros T, Ticconi C, Piccoli E, Zicari A, Piccione E et al. Macrophage migration inhibitory factor in human pregnancy and labor. *Am J Reprod Immunol* 2002;48:404–9.
- [140] Helaakoski T, Vuori K, Myllyla R, Kivirikko KI, Pihlajaniemi T. Molecular cloning of the alpha-subunit of human prolyl 4-hydroxylase: the complete cDNA-derived amino acid sequence and evidence for alternative splicing of RNA transcripts. *Proc Natl Acad Sci U S A* 1989; 86:4392–6.
- [141] Cioffi CL, Qin Liu X, Kosinski PA, Garay M, Bowen BR. Differential regulation of HIF-1 α prolyl-4-hydroxylase genes by hypoxia in human cardiovascular cells. *Biochem Biophys Res Commun* 2003; 303:947–53.
- [142] Hon WC, Wilson MI, Harlos K, Claridge TD, Schofield CJ, Pugh CW et al. Structural basis for the recognition of hydroxyproline in HIF-1 α by pVHL. *Nature* 2002;417:975–8.